Title: A stoichiometric approach to estimate sources of mineral-associated soil organic matter

Authors: Yi Chang¹, Noah W. Sokol², Kees Jan van Groenigen³, Mark. A. Bradford⁴, Dechang Ji¹, Thomas W. Crowther⁵, Chao Liang⁶, Yiqi Luo⁷, Yakov Kuzyakov^{8, 9}, Jingkuan Wang¹, Fan Ding¹

¹ College of Land and Environment, Shenyang Agricultural University, Shenyang, 110866, China
 ² Physical and Life sciences Directorate, Lawrence Livermore National Laboratory, Livermore, CA, USA

³ Department of Geography, College of Life and Environmental Sciences, University of Exeter, Exeter EX4 4 RJ, UK

⁴ Yale School of the Environment, Yale University, New Haven, CT, 06511, USA

⁵ Department of Environmental Systems Science, Institute of Integrative Biology, ETH Zürich, Zürich, Switzerland

⁶ Key laboratory of Forest Ecology and Management, Institute of Applied Ecology, Chinese Academy of Sciences, Shenyang 110016, China

⁷ School of Integrative Plant Science, Cornell University, Ithaca, NY 14853, USA

⁸ Department of Soil Science of Temperate Ecosystems, Department of Agricultural Soil Science, Georg-August University of Göettingen, Göettingen, Germany

⁹ Agro-Technological Institute, Peoples Friendship University of Russia (RUDN University),
117198, Moscow, Russia

Yi Chang and Noah W. Sokol should be considered joint first author. Fan Ding and Jingkuan Wang should be considered joint corresponding author.

Correspondence:

Fan Ding, College of Land and Environment, Shenyang Agricultural University, Shenyang 110866,

China.

Email: dingfan1985@syau.edu.cn

Jingkuan Wang, College of Land and Environment, Shenyang Agricultural University, Shenyang

110866, China.

Email: jkwang@syau.edu.cn

1 ABSTRACT

Mineral-associated soil organic matter (MAOM) is the largest, slowest-cycling pool of carbon (C) in 2 the terrestrial biosphere. MAOM is primarily derived from plant and microbial sources, yet the 3 relative contributions of these two sources to MAOM remain unresolved. Resolving this issue is 4 5 essential for managing and modeling soil carbon responses to environmental change. Microbial biomarkers, particularly amino sugars, are the primary method used to estimate microbial versus 6 plant contributions to MAOM, despite systematic biases associated with these estimates. There is a 7 clear need for independent lines of evidence to help determine the relative importance of plant versus 8 microbial contributions to MAOM. Here, we synthesized 288 datasets of C/N ratios for MAOM, 9 particulate organic matter (POM), and microbial biomass across the soils of forests, grasslands and 10 croplands. Microbial biomass is the source of microbial residues that form MAOM, whereas the 11 POM pool is the direct precursor of plant residues in MAOM. We then used a stoichiometric 12 approach - based on two-pool, isotope-mixing models - to estimate the proportional contribution of 13 plant residue versus microbial sources to the MAOM pool. Depending on the assumptions 14 underlying our approach, microbial inputs accounted for between 34%-47% of the MAOM pool, 15 whereas plant residues contributed 53%-66%. Our results therefore challenge the existing hypothesis 16 that microbial contributions are the dominant constituents of MAOM. We conclude that 17 biogeochemical theory and models should account for multiple pathways of MAOM formation, and 18 that multiple independent lines of evidence are required to resolve where and when plant versus 19 20 microbial contributions are dominant in MAOM formation.

21

3

Keywords: soil carbon, mineral-associated organic matter, plant carbon, particulate organic matter,
 soil organic matter dynamics, meta-analysis

24

25 **INTRODUCTION**

Soil organic matter (SOM) is Earth's largest actively cycling reservoir of carbon (C). Mineral-26 associated organic matter (MAOM) is the largest pool of SOM in Earth's mineral soils (>1500 Pg C). 27 as well as the slowest-cycling, since physico-chemical interactions between SOM and the mineral 28 matrix limit microbial access and decomposition (Kleber et al., 2021; Sokol et al., 2022b). MAOM is 29 primarily composed of relatively simple biomolecules, which interact with soil minerals to form 30 MAOM via two dominant pathways (Sokol et al., 2019). In the first pathway, plant inputs are 31 assimilated by microorganisms and transformed into microbial cellular components; as microbial 32 cells die and turnover, their residues interact with soil minerals to form 'microbial-derived' MAOM 33 (Liang et al., 2017). In the second pathway, plant inputs directly interact with soil minerals without 34 passing through microbial cells (i.e., 'plant-derived MAOM') – either in their intact form (e.g., direct 35 sorption of simple sugars and amino acids from root exudates), or after partial decomposition of 36 more complex plant compounds (e.g., lignin) by extracellular enzymes into simpler compounds 37 (Sokol et al., 2019). 38

While both microbial and plant inputs form MAOM, their relative contributions remain unresolved, as do the environmental factors controlling their relative contributions (<u>Angst et al.</u>, <u>2021</u>; <u>Whalen et al.</u>, <u>2022</u>). Resolving these unknowns will inform (1) our basic understanding of the dominant controls on MAOM formation, and (2) how these controls are represented in

biogeochemical models (Liang et al., 2017; Cotrufo & Lavallee, 2022; Sokol et al., 2022b). For
instance, the formation of 'microbial-derived MAOM' centers microbial traits like growth rate and
carbon-use efficiency (CUE) (Kallenbach et al., 2016; Hu et al., 2022a), whereas the formation of
plant-derived MAOM may center traits like extracellular enzyme production, which can directly
trade-off with growth rate and CUE (Malik et al., 2019; Sokol et al., 2022a).

Several approaches have been used to quantify plant versus microbial contributions to MAOM, 48 each with their own strengths and shortcomings (Whalen et al., 2022). These approaches include: (i) 49 microbial biomarker analysis (e.g., amino sugars, lipids) (Ludwig et al., 2015; Angst et al., 2021), (ii) 50 'molecular fingerprinting' approaches, that compare spectra or peaks of soil C to plant or microbial 51 inputs (e.g., NMR or pyrolysis GC-MS) (Simpson et al., 2007; Grandy & Neff, 2008), and (iii) 52 mathematical models (Klink et al., 2022). To date, the most common approach to quantify the 53 contribution of microbial products to SOM are amino sugar microbial biomarkers (Appuhn & 54 Joergensen, 2006; Liang et al., 2020). While amino sugars biomarkers have shed light on the relative 55 contributions of microbial compounds to SOM and MAOM pools, the amino sugar approach also 56 comes with several limitations (Joergensen, 2018; Liang et al., 2019; Whalen et al., 2022). Due to 57 limitations in the amino sugar biomarker approach and other existing approaches, there is a critical 58 need for additional, new approaches to independently and quantitatively determine plant versus 59 microbial contributions to MAOM (Liang et al., 2020; Whalen et al., 2022). As each existing and 60 new approach will likely bring its own unique set of shortcomings, multiple independent lines of 61 evidence are required to develop the most robust estimates. 62

63 The stoichiometry of microorganisms and plants provides one such additional and independent 64 source of data for evaluating the relative contributions of plant and microbial inputs to MAOM

formation. SOM consistently has a larger C/N than microbial biomass C/N and a lower C/N than 65 plant inputs because it contains a mixture of plant and microbial residues (Coonan et al., 2020). By 66 extension, if the C/N ratio of MAOM is intermediate between microbial biomass and plant residues, 67 MAOM then also contains some proportion of the two. Comparing C/N ratios of these soil C pools 68 may yield important and independent insights on the contributions of plant and microbial residues to 69 MAOM pools. To date, few if any studies have comprehensively compared the C/N ratios of 70 microbial biomass, plant residues and MAOM across a broad range of environmental conditions, nor 71 used their stoichiometry to estimate plant residue versus microbial contributions to MAOM. 72

Here, we put forward a new line of evidence to estimate the contributions of plant and microbial 73 residues to MAOM based on their stoichiometry. Plant residues are primarily incorporated into 74 MAOM via the particulate organic matter (POM) pool (Coonan et al., 2020; Witzgall et al., 2021; 75 Cotrufo & Lavallee, 2022), which consists largely of partially decomposed plant compounds (Guigue 76 et al., 2021). Thus, the C/N ratio of POM can be used to estimate plant residue inputs to MAOM. 77 Central to the isotope mixing approach is that the values of the source materials, such as POM and 78 microbial biomass, differ to the mixture (i.e., MAOM) and to one another, bracketing the mixture. 79 These differences generally hold for POM, MAOM and microbial biomass (Cleveland & Liptzin, 80 2007; Xu et al., 2013; Cotrufo et al., 2019; Amorim et al., 2022). Although the stoichiometry of these 81 pools is commonly expressed as C/N ratios, we used the fractional abundance of N (i.e., N/(C+N)) to 82 estimate their relative contributions. This approach avoids the spurious inferences that can arise from 83 using ratios (e.g., Jasieński & Bazzaz, 1999), and instead adapts the approach used in enriched 84 isotope mixing-models in ecology to identify the contributions of sources to a mixture. Specifically, 85 as with enriched stable isotopic approaches, ratios cannot be used in mixing equations because the 86

87	difference between the light and heavy isotope in the denominator is no longer essentially a constant
88	- which it is with natural abundance approaches - meaning that ratios must be converted to fractional
89	abundances for mixing equations (Fry, 2006). We therefore propose that the fractional abundance of
90	N in microbial biomass and POM can be used as end-members in a mixing model to differentiate
91	their contributions to MAOM, providing a novel line of evidence to help evaluate the relative
92	contributions of plant- and microbial-derived materials to MAOM.

We collected 288 published sets of data that report C/N ratios of microbial biomass, POM and MAOM. These data mainly cover three ecosystems (forests, grasslands, croplands) and different soil depths. We expected that MAOM would have higher C/N values than microbial biomass and lower C/N values than POM, because MAOM typically contains a mix of both plant and microbial residues. Subsequently, we used the ecosystem-specific combination of the fractional abundance of N, i.e., [N/(C+N)] of POM, microbial biomass, and MAOM to assess the relative plant and microbial residue contributions to MAOM.

100

101 **METHODS**

102 Data collection and extraction

We collected data from peer-reviewed articles published before August 2022 on Google Scholar and Web of Science, which simultaneously reported the C/N ratios (or carbon and nitrogen contents) of three soil organic matter pools: 1) microbial biomass, 2) POM (or light fraction), and 3) MAOM (or heavy fraction). MAOM and POM pools were defined both by size fractionation and density fractionation, as has been done in a previous meta-analysis (Georgiou et al., 2022). For each study in our dataset, we also extracted corresponding data on climate (mean annual temperature (MAT), mean

annual precipitation (MAP)) and soil properties (clay content, pH, soil organic carbon, total N, soil C/N). The missing data of climate and soil properties were supplemented by searching from global GIS datasets by latitude and longitude. These variables were selected to represent the broadest variation in environmental characteristics following <u>van den Hoogen et al. (2019)</u>, and to capture the environmental drivers that have been hypothesized to influence microbial growth and activity in soil (Crowther et al., 2019).

Papers had to meet the following criteria to be included in our dataset: microbial biomass 115 carbon (MBC) and nitrogen (MBN) were simultaneously measured by the chloroform fumigation 116 extraction method. This is because separate measurements (e.g., MBC as measured by fumigation-117 incubation method and MBN by steam distillation for mineral N) of the two indexes would increase 118 the uncertainty of microbial C/N. To keep consistency among all studies with various fractionation 119 methods, multiple sub-POM fractions, e.g., fine POM, coarse POM, free POM, or occluded POM in 120 some studies, were unified to obtain an overall C/N of POM by dividing the combined total C 121 content of these fractions by their total N content. 122

In total, 294 groups of C/N ratios (one group included one C/N ratio of microbial biomass, POM, and MAOM, respectively) were collected from 36 peer-reviewed articles. Improbably low C/N values – based on empirical and theoretical knowledge – were removed (i.e., one group of POM C/N that was 1.9, and five groups of Microbial C/N that were < 2), resulting in 288 groups of C/N ratios from 36 peer-reviewed articles. All C/N data of POM, microbial biomass, and MAOM in this research are available at https://doi.org/10.5281/zenodo.10147884.

129

130 *Calculations and statistical analysis*

A paired sample *t*-test was used to test the pairwise differences in C/N ratios between microbial biomass and MAOM, and between POM and MAOM, respectively. We used the combination of the fractional abundance of N of POM and microbial biomass to assess the relative contributions of plant and microbial residues to MAOM (Figure 1). We assumed that microbial biomass C/N ratio was similar to microbial necromass C/N, so the fractional abundance of N was the same between the two pools. The fractional abundance of N, i.e., [N/(C+N)], in microbe, POM, and MAOM pools was calculated based on their corresponding C/N ratios as following:

$$[N/(C+N)] = 1/(1+C/N)$$
(1)

We used the fractional abundance of N in the two-end-member model to differentiate the contributions of microbial and plant residues to MAOM following:

$$f \times [N/(C+N)]_{Microbe} + (1-f) \times [N/(C+N)]_{POM} = [N/(C+N)]_{MAOM}$$
(2)

Where f is the proportion in MAOM that is derived from microbial residues, and thus 1-f is the fraction in MAOM originating from plant residues (i.e., POM). [N/(C+N)]_{Microbe} is the fractional abundance of N in microbial biomass. [N/(C+N)]_{POM} is the fractional abundance of N in POM. [N/(C+N)]_{MAOM} represents the fractional abundance of N in MAOM.

144 Therefore, the proportion of microbial residue to MAOM (f) can be estimated as:

$$f = \frac{[N/(C+N)]_{MAOM} - [N/(C+N)]_{POM}}{[N/(C+N)]_{Microbe} - [N/(C+N)]_{POM}}$$
(3)

145 When $[N/(C+N)]_{MAOM} > [N/(C+N)]_{Microbe}$ (i.e., $C/N_{MAOM} < C/N_{Microbe}$), the values of f are larger 146 than 1; when $[N/(C+N)]_{MAOM} < [N/(C+N)]_{POM}$ (i.e., $C/N_{MAOM} > C/N_{POM}$), f is smaller than 0. Within

our collected 288 groups of data, 70 groups had C/N_{MAOM} < C/N_{Microbe}, and 39 groups had 147 $C/N_{MAOM} > C/N_{POM}$. $C/N_{MAOM} < C/N_{Microbe}$ may partly be attributed to MAOM containing some 148 proportion of mineral N besides organic matter, e.g., the adsorption of ammonium on soil minerals 149 (Mortland, 1959; Adams Jr. & Stevenson, 1964). However, according to Bimüller et al. (2014), the 150 proportion of mineral N to total N in silt or clay sized MAOM was only 0.6~2.5%, thereby not 151 substantially changing C/N ratios of MAOM. Accordingly, we speculated that C/N_{MAOM} < C/N_{Microbe} 152 indicated that 100% of MAOM is from microbial residues and $C/N_{MAOM} > C/N_{POM}$ indicates 100% 153 of MAOM is derived from plant residues and nil derived from microbial residues, and the difference 154 resulted from the measurement error of C/N. Therefore, these abnormal f-values were replaced by 1 155 and 0, respectively. We analyze and present the data both excluding and including these abnormal 156 points. 157

The C/N ratios of organic matter pools and the proportion of microbial residues in MAOM were compared among grasslands, forests, and croplands using one-way ANOVA. As the sampled soil depth differed between experiments, the depth was expressed as the arithmetic mean of the upper and lower boundaries of each layer. Soil arithmetic mean depths were divided into two categories: < 20 cm as topsoil and \geq 20 cm as subsoil; we then compared the average proportion of microbial residues in MAOM between the two depth categories using one-way ANOVA.

Regression analysis was performed to look at correlations among the C/N ratios of MAOM and microorganisms, and between MAOM and POM. In addition, random forest analysis was used to quantify the relative explanatory power of climate (MAT and MAP) and soil properties (clay content, pH, soil organic carbon, total N, soil C/N) on the microbial contribution to MAOM. All the paired sample *t*-tests, regression analyses and ANOVA analyses were conducted in SPSS 25.0 (SPSS Inc.,

Chicago, IL, USA). Random forest analysis was performed using the R "randomForest" package and
the "rfPermute" package in the R Statistical Environment (Version 4.1.0, R Core Team).

171

172 Sensitivity test under scenarios for contributions of plant DOM to plant-derived MAOM

Root exudation and DOM from leaf litter leachate are also potential contributors to MAOM (Cotrufo 173 & Lavallee, 2022), thus plant DOM may affect the estimation of our two-end-member model. If the 174 C/N ratio of plant DOM is close to that of POM, the incorporation of DOM in the above two-end-175 member model does not substantially change the contribution of microbial residues to MAOM. 176 However, some N-poor compounds (e.g., carbohydrates or aromatic acids) also likely contribute to 177 MAOM by sorption with reactive mineral phases (Kramer et al., 2012; Whalen et al., 2022). 178 According to Qualls and Haines (1991), C/N ratios of N-poor hydrophobic acids in DOM across 179 180 various soil horizons and in streams ranged from 34 to 73. In this study, we assumed a C/N ratio of 50 for these N-poor components in DOM (the fractional abundance of N thereby 1/51), to test the 181 sensitivity of our approach to the inclusion of plant DOM in MAOM. Unfortunately, no studies 182 reported the proportion of DOM in plant-derived MAOM (Whalen et al., 2022). We assumed two 183 scenarios, where the proportion of plant DOM contributions to plant-derived MAOM were either low 184 (i.e., 10%) or high (i.e., 50%). We then adjusted the fractional abundance of N of plant-derived 185 organic matter in these scenarios as follows: 186

Scenario 1:
$$[N/(C+N)]_{POM+DOM} = [N/(C+N)]_{POM} \times 90\% + [N/(C+N)]_{DOM} \times 10\%$$
 (4)

Scenario 2:
$$[N/(C+N)]_{POM+DOM} = [N/(C+N)]_{POM} \times 50\% + [N/(C+N)]_{DOM} \times 50\%$$
 (5)

187 Where $[N/(C+N)]_{DOM}$ is assumed to be 1/51. Then, we used the new combined 188 $[N/(C+N)]_{POM+DOM}$ instead of $[N/(C+N)]_{POM}$ to estimate the microbial contribution to MAOM using 189 equation (1).

190

191 Sensitivity test under scenarios for proportions of microbial residues in POM

Although POM is primarily composed of partially decomposed plant fragments (<u>Golchin et al., 1994</u>;
 Lavallee et al., 2020; Guigue et al., 2021), isotopic evidence suggests that some microbial necromass

194 exists as POM (Wang et al., 2020), such as larger fungal fragments (Lavallee et al., 2020). Similarly,

195 small amounts of microbial-derived amino sugars have been measured in POM (<u>Turrión et al., 2002</u>; 196 <u>Griepentrog et al., 2014</u>). Because our approach uses POM to represent the contribution of plant 197 residue inputs to MAOM, microbial residues in the POM fraction would decrease the C/N ratio and 198 increase the fractional abundance of N, and may lead to an overestimation of the plant contribution 199 to MAOM (an underestimation of the microbial contribution).

To assess the possible impact of this uncertainty, we estimated the proportion of microbial 200 residue in POM, based on the amino sugar data in Griepentrog et al. (2014) and using the conversion 201 method in Wang et al. (2021). These results suggest that the total fungal and bacterial necromass C 202 accounted for 10% in free light fraction C and 15% in occluded light fraction C (light fraction is 203 equivalent to POM, Table S1). Based on these results, we ran a simulation under different scenarios 204 of microbial residue proportions in POM to evaluate how sensitive our assumptions were to the 205 possibility that POM contains some microbial biomass and/or necromass. We assumed: 1) that if the 206 proportion of microbial residues in POM is f_1 , then the proportion of plant residues in POM is 1 - f_1 ; 207 and 2) that if the proportion of microbial residues in MAOM is f₂, then the proportion of plant 208

- residues in MAOM is $1 f_2$. Accordingly, the relationships between the fractional abundances of N in
- 210 microbial biomass, POM and MAOM can be described by the following equations:

$$f_1 \times [N/(C+N)]_{Microbe} + (1-f_1) \times [N/(C+N)]_{Plant} = [N/(C+N)]_{POM}$$
(6)

$$f_2 \times [N/(C+N)]_{Microbe} + (1-f_2) \times [N/(C+N)]_{Plant} = [N/(C+N)]_{MAOM}$$
(7)

211 Where $[N/(C+N)]_{Plant}$ represents the fractional abundance of N in plant residue in POM and 212 MAOM. Combining equations (6) and (7), we arrived at the following equation:

$$f_{2} = \frac{[N/(C+N)]_{MAOM} - [N/(C+N)]_{Microbe}}{[N/(C+N)]_{POM} - [N/(C+N)]_{Microbe}} \times f_{1} + \frac{[N/(C+N)]_{POM} - [N/(C+N)]_{MAOM}}{[N/(C+N)]_{POM} - [N/(C+N)]_{Microbe}}$$
(8)

We assumed two scenarios of when f_1 was 10% and 15%, as described above. Finally, we can obtain the proportion of microbial residues in MAOM (f_2) under the two scenarios, based on the N/(C+N) values of MAOM, POM, and microbe pools for a given soil.

216

217 **RESULTS**

218 *C/N ratios of organic matter pools*

219 The mean C/N ratio of MAOM was higher than that of microbial biomass in forests (p < 0.001),

croplands (p < 0.05), and grasslands (p = 0.11), as well as across all ecosystem types (p < 0.001)

- 221 (Figure 2). The C/N ratio of MAOM was lower than that of POM in all the three types of ecosystems
- (all p < 0.001) (Figure 2). The fact that the C/N ratio of MAOM fell between the C/N ratio of POM
- and microbial biomass (Figure 2) supports theoretical expectations that microbial and plant residues
- are both contributors to MAOM.
- POM had a larger mean C/N ratio in forests (24.4) than in grasslands (19.2) and croplands (15.5)

(p < 0.001, Figure 2). MAOM C/N was also larger in forests (13.8) than grasslands (12.4) and croplands (11.0) (p < 0.001). The mean C/N ratios of microbial biomass were, by contrast, approximately the same across forests, grasslands, and croplands (p = 0.30). Despite the differences of the mean C/N ratios of the SOM fractions and similarities microbial biomass among the three ecosystem types, the C/N ratios varied markedly within each ecosystem type. Overall, MAOM fractions had a narrower range (from 7.1 to 41.5) than those of POM (7.9 to 104.6) and microbial biomass (from 2.1 to 37.1).

MAOM C/N was positively associated with microbial biomass C/N in forests and grasslands (p 233 < 0.01, Figure 3b, and 3c), as well as across all ecosystem types (p < 0.05, Figure 3a), but not in 234 croplands (Figure 3d). The slope of the regression line between MAOM and microbial biomass C/N 235 was steeper in forests (0.29) than in grasslands (0.14) and was also steeper than the slope across the 236 entire data set (0.09). MAOM C/N was positively associated with POM C/N across all ecosystem 237 types (p < 0.001, Figure 3a), within forests (p < 0.001, Figure 3b) and croplands (p < 0.05, Figure 238 3d), but not within grasslands (Figure 3c). The slope of the regression line between MAOM and 239 POM C/N was steeper in forests (0.15) and across the entire data (0.12) than in croplands (0.05). 240

241

242 Microbial contributions to MAOM depend on ecosystem and soil depth

Stoichiometric analysis based on fractional abundances estimated that, on average, microbial inputs accounted for ~34% of contributions to MAOM across all environmental contexts (Figure 4a), whereas plant residue inputs (via POM) accounted for ~66%. The microbial contribution to MAOM was higher in forests (38%) and croplands (36%) than in grasslands (27%) (p = 0.07, Figure 4a). Across all ecosystem types, the microbial contribution to MAOM increased by 1.3 times from 33%

in topsoil (< 20 cm) to 42% in subsoil (\geq 20 cm) (p = 0.09, Figure 4b). Unfortunately, we did not 248 have enough data to estimate the effect sizes of causal variables that might be generating the 249 differences between soil depths and ecosystems. We instead used random forest analysis to look at 250 which variables explained the most variation in the estimated contributions to MAOM, which 251 suggested that soil clay content was the most important factor influencing the microbial contribution 252 to MAOM (Figure 5). Microbial contribution to MAOM positively correlated with soil clay content 253 (p < 0.001), Figure S1a), suggesting that the importance of microbial contributions to MAOM may be 254 particularly pronounced in clay and loamy soils. 255

We observed a larger microbial contribution to MAOM on samples isolated via density 256 fractionation than via size fractionation, though the general patterns observed (e.g., microbial 257 contribution less than 50%, and lower microbial contribution in grassland than other ecosystem 258 types) were consistent between both methods (Figure S2). Density fractionation is generally 259 considered to be more effective at isolating the mostly plant-derived, particulate organic matter 260 fraction (Leuthold et al., 2023), which may explain why this method was associated with more 261 microbial-derived MAOM (Figure S2). Since there were more samples in our dataset isolated via 262 density fractionation versus size fractionation (n=99 vs. n=80), this may have dampened the 263 magnitude of effect we observed in terms of the overall plant contributions to MAOM. 264

265

266 *Sensitivity analysis*

In addition to POM, another potentially important source of plant C to the MAOM pool is DOM. We used two illustrative scenarios to estimate how N-poor DOM (assuming a C/N ratio of 50) inputs may affect our model results – 10% and 50% contributions of DOM to the plant-derived MAOM

pool (see Methods). When the DOM contribution increases, the N fractional abundance of the 270 combined POM+DOM pools decreases (i.e., a C/N increase) (Eq. 4 and Eq. 5), which then leads to 271 estimates from the mixing models of increased microbial contributions to MAOM (Eq. 3). When 272 assuming the proportion of DOM was 10%, the average microbial contributions were 35% (Figure 273 S3a) – which is relatively close to our estimate when DOM was excluded (i.e., ~34%, Figure 4a). 274 When assuming the proportion of DOM was 50%, the average microbial contribution increased to 275 47% (Figure S3c) – still less than 50% of the total pool. The microbial contribution to MAOM was 276 higher in subsoil than in topsoil, regardless of whether the DOM contribution to plant-derived 277 MAOM was 10% (Figure S3b) or 50% (Figure S3d). 278

Although POM is primarily composed of plant residues, it may contain some microbial residues (Guigue et al., 2021). To reveal the uncertainty associated with microbial contributions via POM, we modeled the two scenarios of 10% and 15% microbial residues in POM, based on the data in Griepentrog et al. (2014), as shown in Table S1. The average microbial contributions to MAOM increased from 34% to 41% and 44% under the two scenarios, respectively (Figure S4a,c). The comparisons of microbial contributions among different ecosystems and soil depths (Figure S4) and its relationship with climate and soil properties (Figure S5), did not substantially change.

Finally, in our dataset, there were dozens of datapoints where $C/N_{MAOM} < C/N_{Microbe}$, or $C/N_{MAOM} > C/N_{POM}$, which – using the mixing models – then led to estimated microbial contributions to MAOM of 100% and 0, respectively (see Methods). Instead of eliminating these datapoints from our analysis, we ran our analyses both without these datapoints (i.e., model results described above (Figure 4)), as well as with these abnormal data points (Figure S6), to see how they affected our results. When they were included, the average microbial contribution increased from

34% to 45%, i.e., still lower than the plant contributions (Figures 4a and S6a). The average microbial
contribution increased from 38% to 43% in forests, from 27% to 46% in grasslands, and from 36% to
46% in croplands (Figures 4a and S6a). The higher microbial contribution to MAOM in subsoil than
in topsoil and the relative explanatory power of climate and soil factors for the microbial
contributions to MAOM did not change (Figures S6b and S7).

297

298 **DISCUSSION**

There is an increasingly popular view that microbial residues strongly dominate MAOM pools 299 (Bradford et al., 2013; Cotrufo et al., 2013; Kögel-Knabner, 2017; Oldfield et al., 2018; Creamer et 300 al., 2019; Buckeridge et al., 2020; Liang et al., 2020; Cotrufo & Lavallee, 2022; See et al., 2022). 301 This view is built both on qualitative and semi-quantitative sources of evidence, such as imaging 302 techniques that show microbial necromass-mineral associations (e.g., scanning electron microscopy; 303 Miltner et al., 2012), as well as more directly quantitative approaches, such as the upscaling of 304 microbial biomarkers (e.g., amino sugars) (Liang et al., 2019; Wang et al., 2021), 'molecular 305 fingerprinting' approaches (e.g., ¹³C-NMR or py-GC-MS) (Lehmann et al., 2007; Grandy & Neff, 306 2008; Lehmann et al., 2008; Solomon et al., 2012), and mathematical models (e.g., Bayesian 307 Inference isotopic mixing model based on ¹³C and ¹⁵N natural abundance) (Klink et al., 2022). Yet, 308 caveats in these existing approaches introduce uncertainty into quantitative estimates of microbial 309 versus plant contributions to MAOM. This has precipitated calls for new, independent lines of 310 evidence (Whalen et al., 2022). Here, to address that call, we conducted a meta-analysis of C/N 311 ratios of MAOM, microbial biomass, and POM across different environmental contexts, and used a 312 novel stoichiometric approach (via the fractional abundance of N) to estimate the contributions of 313

314 plant residues (via the POM pool) and microbial residues to MAOM. Below, we first discuss the 315 results of the meta-analysis, followed by the results of our stoichiometric approach.

Our meta-analysis showed that MAOM C/N ratios are greater than microbial C/N ratios and 316 lower than POM C/N ratios within each of the three ecosystem types, as well as across all ecosystem 317 types (Figure 2). These findings suggest that MAOM contains both microbial and plant residues, in 318 agreement with prior studies. For example, a meta-analysis based on 74 studies across 29 countries 319 showed that sand-sized fractions (equivalent of POM) have larger C/N ratios (median of 16.7) than 320 silt-sized (median of 13.1) and clay-sized MAOM fractions (median of 9.6) (Amorim et al., 2022). 321 This trend was also observed in a forest and a grassland soil (Ding et al., 2014) and two cropland 322 soils including upland and paddy (Ding et al., 2018). An analysis based on 9,415 data points of 323 European forest and grassland soils reported that the C/N ratio of MAOM (13 \pm 5) was lower and 324 less variable than that of POM (22 ± 15) (Cotrufo et al., 2019). Moreover, we found a close link 325 between MAOM and POM across the entire dataset, especially in forest and cropland soils (Figure 326 3). Similar close relationships between C/N ratios of POM and MAOM, as well as for δ^{13} C, C–H 327 (aliphatic)/C=O, and C=C (aromatic)/C=O), were observed across 156 soils spanning diverse 328 ecosystems (tundra to tropics) in North America (Yu et al., 2022). Based on these relationships, they 329 concluded that POM and MAOM are coupled, and that plant residues contribute substantially to the 330 MAOM pool. 331

Overall, the results from our stoichiometric approach suggest that microbial inputs form less than 50% of MAOM across different ecosystem contexts. The overall mean microbial contribution was 34%-47% (Figures 4 and S3c), whereas the overall mean plant residue contribution to MAOM was 53%-66%. Our results align with other studies, which have also suggested that microbial inputs

may not be the dominant constituent of the MAOM. For instance, in a synthesis of microbial 336 biomarker studies, Angst et al. (2021) estimated an average contribution of microbial necromass to 337 MAOM ("silt and clay fractions" in their study) of 39%, based on 60 groups of amino sugars data. 338 Similarly, based on amino sugar data, microbial contributions to MAOM were 21%-30%, in an 339 organic grain crop-cover crop rotation field in the USA (Zhang et al., 2022), and 13%-19% in 340 subtropical forest and cropland in the southwestern karst region of China (Hu et al., 2022b). 341 Chemical composition-based approaches have also suggested microbial inputs may not be dominant. 342 For example, a study using pyrolysis-field ionization mass spectrometry, which used lipid and 343 carbohydrates as biomarkers in four arable soils of widely differing properties, showed a 1:1 344 contribution for microbial and plant compounds to MAOM (Ludwig et al., 2015). A study using ¹³C-345 NMR on an acidic forest soil concluded that microbial contributions to total SOM were less than 5% 346 347 (Simpson et al., 2007). As emphasized in a recent review, the estimated proportions of MAOM that are microbial versus plant derived are method- and context-dependent (see Whalen et al. (2022) for a 348 discussion on how different methods may influence these estimates). 349

We found that the contribution of microbial residues to MAOM increased with soil depth (Figure 350 4b). This is consistent with previous studies showing that the contribution of amino sugars to the 351 SOM pool was greater in subsoil (Liang & Balser, 2008; Angst et al., 2018). These patterns were 352 suggested to be caused by the different distributions across soil depths of plant and microbial 353 residues. Kaiser et al. (2004) observed that the concentration of plant derived DOC (e.g., lignin-354 derived phenols) decreased with depth while that of microbial derived DOC (e.g., amino sugars) 355 increased or remained constant. Consistent with this observation, we found a larger proportion of 356 plant residues (indicated by POC in total SOC) in topsoil than in subsoil (p < 0.001, Figure S8). 357

Kaiser et al. (2004) explained that the decrease in lignin-derived phenols was due to their strong affinity with Al and Fe oxides-hydroxides (Kaiser et al., 2002), and that hydrophobic DOC that was preferentially retained when transported through the mineral soil contained few amino sugars (Kaiser et al., 2001). Another explanation could be that the major dissolved molecules from plants are coming from the soil surface; in contrast, the microbial transformations are "similar" in top- and subsoil.

Among soil properties, texture was the most important factor influencing the relative 364 contribution of microbial residues to MAOM (Figure 5), which increased with increasing clay 365 content (Figure S1a). These results suggest that clay and loamy soils generally have larger microbial 366 contributions to MAOM than sandy soils. This observation is supported by Yu et al. (2022) who 367 found that the difference between C/N ratios of POM and MAOM increased with silt and clay 368 contents (the larger the difference, the smaller the plant contribution to MAOM, and thereby the 369 larger the microbial contribution). Similarly, the abundance of amino sugars in global grassland soils 370 increases with clay contents (Ma et al., 2018). Compared to sand and silt particles, clay particles 371 have much larger negative charged mineral surfaces and stronger aggregation (Ding et al., 2014; 372 Ding et al., 2018), causing a stronger affinity for microbe-derived molecules (e.g., amino acids and 373 amino sugars, containing the positively charged -NH₂ groups). These properties can protect the 374 molecules against re-utilization by other microbes or higher trophic groups (Elliott et al., 1980; Ding 375 et al., 2015; Islam et al., 2022) and enzymatic attacks (Baldock & Skjemstad, 2000). Accordingly, 376 Grandy and Neff (2008) reported that microbially-derived compounds (e.g., N-containing 377 compounds, lipids, waxes, aliphatics, and carbohydrates) increase with decreasing soil particle size. 378 In addition, soils with high clay contents commonly have higher nutrient contents (Ristori, 1979), 379

380 which favor the growth and turnover of microorganisms, as well as the formation of microbial 381 residues (<u>Creamer et al., 2016</u>).

Notably, the contribution of microbial residues to MAOM were similar between forests and 382 croplands, but they were higher than that in grasslands (p = 0.07, Figure 4a). These results likely 383 reflect that within our dataset, clay content in grasslands was lower than in croplands and forests 384 (Figure S1b). Our results are in line with Hu et al. (2022b) who observed similar microbial 385 contribution to MAOM C in forest and cropland soils for both bacteria and fungi. In contrast, Angst 386 et al. (2021) found that grasslands favor microbial necromass accumulation in MAOM relative to 387 croplands or forests, however their grassland data points were very limited (n = 2). Importantly, the 388 contribution of microbial residues to MAOM does not mirror its contribution to bulk SOM (Angst et 389 al., 2021). Two recent meta-analysis studies, both of which used the amino sugars biomarker 390 approach, found that the proportion of microbial necromass C in bulk SOC was lower in forests than 391 grasslands and croplands (Liang et al., 2019; Wang et al., 2021). This could be due to the greater 392 dominance of POM (mainly plant residues) in forest SOM pools (Figure S8), a result consistent with 393 a recent multi-site analysis (Cotrufo et al., 2019). 394

There are important limitations to our method, which should be addressed by future studies to help build upon and refine our approach. First, our approach does not account for the possible impact of microbial death pathways on the chemical composition of microbial necromass (<u>Camenzind et al.</u>, <u>2023</u>). Once data becomes available on the impact of microbial death pathways on necromass chemistry, it should be used to fine-tune our approach. Second, other sources of organic input may be important for MAOM formation in addition to POM and microbial body residues, such as plantderived DOM, amendments in croplands, or microbial extracellular products. To date, minimal data

exists quantifying the role of these inputs on MAOM formation. Moreover, there is limited data for 402 what proportion of POM may be derived from microbial inputs (Lavallee et al., 2020). It will be 403 critical for future empirical efforts to determine if other sources of C input may influence the relative 404 contributions of plant versus microbial inputs in different ecosystem contexts, to better constrain the 405 proportion of microbial residues in the POM pool, as well as more directly ascertain how plant DOM 406 and microbial extracellular residues contribute to the MAOM pool (Whalen et al., 2022). Third, our 407 uncertainty estimates did not include measurement errors of C/N ratios, as data errors were not 408 always reported. We encourage future studies to clearly report errors, make data openly available, 409 and to use statistical approaches (such as Bayesian mixing models) that carry such errors forward. 410 Last, few if any datasets exist which use multiple approaches – on a common set of soil samples – to 411 measure plant versus microbial contributions to MAOM. As all current methods come with their own 412 413 unique limitations, it will be critical to design studies that employ a suite of complementary approaches (such as different biomarkers, isotopic data, C/N ratios, chemical composition data, etc.) 414 to develop the most robust estimates of plant versus microbial contributions to MAOM. 415

416

417 CONCLUSIONS

Our global comparison of the C/N ratios of microbial biomass, POM, and MAOM in forests, grasslands, and croplands demonstrated that MAOM C/N ratios are intermediate between those of POM and microbial biomass. By leveraging these 288 datasets of C/N ratios and using a novel stoichiometric approach (via the fractional abundance of N) to estimate the proportional plant and microbial contributions to the MAOM pool, we found that plant residue contributions to the MAOM pool (53%-66%) exceeded microbial contributions (34%-47%) across ecosystems. These results held

when addressing key potential caveats, such as the role of a third source of input, the role of plant 424 DOM in supplying the MAOM pool, and when including abnormal datapoints that we had removed 425 from our primary analysis. While our results underscore that microbial residues are important 426 constituents of MAOM, especially in deeper soils and in high clay soils, they challenge the 427 increasingly popular view that microbial contributions are the dominant component of MAOM. Our 428 results also underscore the need for other independent lines of evidence that quantitatively 429 differentiate the role of plant versus microbial contributions to the MAOM pool. While there are 430 limitations inherent to each method, multiple lines of evidence will be critical to determine the 431 relative importance of plant versus microbial inputs. Such understanding is needed to advance a 432 more robust theoretical knowledge of SOM dynamics, and for projecting their response to a 433 changing climate. 434

435

436 AUTHOR CONTRIBUTIONS

Yi Chang: Formal analysis, Investigation, Validation, Visualization, Writing – original draft. Noah 437 W. Sokol: Methodology, Validation, Visualization, Writing – original draft, Writing - Review & 438 Editing. Kees Jan van Groenigen: Validation, Writing - Review & Editing. Mark. A. Bradford: 439 Methodology, Validation, Writing - Review & Editing. Dechang Ji: Formal analysis. Thomas W. 440 Crowther: Writing - Review & Editing. Chao Liang: Writing - Review & Editing. Yigi Luo: 441 Writing - Review & Editing. Yakov Kuzyakov: Visualization, Writing - Review & Editing. 442 Jingkuan Wang: Project administration, Resources, Supervision. Fan Ding: Conceptualization, 443 Methodology, Resources, Supervision, Validation, Visualization, Writing - original draft, Writing -444 Review & Editing. 445

446

447 **CONFLICT OF INTEREST STATEMENT**

- 448 The authors declare no competing interests.
- 449

450 DATA AVAILABILITY STATEMENT

The data that support the findings of this study are openly available in Supporting Information and
Zenodo at https://doi.org/10.5281/zenodo.10147884.

453

454 ACKNOWLEDGEMENTS

This work was supported by the National Key Research and Development Plan Project of China 455 (2021YFD1500200), the National Natural Science Foundation of China (42071069 and 32241037), 456 457 and the RUDN University Strategic Academic Leadership Program. Work at Lawrence Livermore National Laboratory by N.W.S. was performed under the auspices of the U.S. DOE OBER, under 458 contract DE-AC52-07NA27344 award #SCW1632. K.J.v.G. was funded by the National 459 Environmental Research Council (NE/W001691/1). We thank Dr. Gerrit Angst in Biology Centre of 460 the Czech Academy of Sciences and Dr. Steven J. Hall in Iowa State University for their valuable 461 comments and suggestions on our paper and Dr. Johan van den Hoogen in Institute of Integrative 462 Biology in ETH Zürich for his help in supplementing the missing environmental factor data in our 463 paper. We also thank three anonymous reviewers for their helpful comments and suggestions. 464

465

466 SUPPLEMENTARY INFORMATION

467 Supplementary data to this article can be found online.

- 468
- 469 **ORCID**
- 470 *Yi Chang* https://orcid.org/0000-0001-8733-7498
- 471 Noah W. Sokol https://orcid.org/0000-0003-0239-1976
- 472 Kees Jan van Groenigen https://orcid.org/0000-0002-9165-3925
- 473 Mark. A. Bradford https://orcid.org/0000-0002-2022-8331
- 474 Dechang Ji https://orcid.org/0000-0001-9280-8864
- 475 Thomas W. Crowther https://orcid.org/0000-0001-5674-8913
- 476 Chao Liang https://orcid.org/0000-0002-9089-6546
- 477 Yiqi Luo https://orcid.org/0000-0002-4556-0218
- 478 Yakov Kuzyakov https://orcid.org/0000-0002-9863-8461
- 479 Jingkuan Wang https://orcid.org/0000-0002-4008-7433
- 480 *Fan Ding* https://orcid.org/0000-0002-7938-1015
- 481

482 **REFERENCES**

- 483 Adams Jr., R. S., & Stevenson, F. J. (1964). Ammonium sorption and release from rocks and 484 minerals. *Soil Science Society of America Journal*, 28 (3), 345-351.
- 485 https://doi.org/10.2136/sssaj1964.03615995002800030014x
- Amorim, H. C., Hurtarte, L. C., Souza, I. F., & Zinn, Y. L. (2022). C:N ratios of bulk soils and
 particle-size fractions: Global trends and major drivers. *Geoderma*, 425, 116026.
 https://doi.org/10.1016/j.geoderma.2022.116026

- Angst, G., Messinger, J., Greiner, M., Häusler, W., Hertel, D., Kirfel, K., . . . Mueller, C. W. (2018).
- 490 Soil organic carbon stocks in topsoil and subsoil controlled by parent material, carbon input
- 491 in the rhizosphere, and microbial-derived compounds. Soil Biology and Biochemistry, 122,
- 492 19-30. <u>https://doi.org/10.1016/j.soilbio.2018.03.026</u>
- Angst, G., Mueller, K. E., Nierop, K. G. J., & Simpson, M. J. (2021). Plant- or microbial-derived? A
 review on the molecular composition of stabilized soil organic matter. *Soil Biology and Biochemistry*, 156, 108189. <u>https://doi.org/10.1016/j.soilbio.2021.108189</u>
- Appuhn, A., & Joergensen, R. G. (2006). Microbial colonisation of roots as a function of plant
 species. Soil Biology and Biochemistry, 38 (5), 1040-1051.
 https://doi.org/10.1016/j.soilbio.2005.09.002
- Baldock, J. A., & Skjemstad, J. O. (2000). Role of the soil matrix and minerals in protecting natural
 organic materials against biological attack. *Organic Geochemistry*, *31* (7), 697-710.
 https://doi.org/10.1016/S0146-6380(00)00049-8
- 502 Bimüller, C., Mueller, C. W., von Lützow, M., Kreyling, O., Kölbl, A., Haug, S., ... Kögel-Knabner,
- 503 I. (2014). Decoupled carbon and nitrogen mineralization in soil particle size fractions of a
- 504 forest topsoil. *Soil Biology and Biochemistry*, 78, 263-273. 505 <u>https://doi.org/10.1016/j.soilbio.2014.08.001</u>
- Bradford, M. A., Keiser, A. D., Davies, C. A., Mersmann, C. A., & Strickland, M. S. (2013).
 Empirical evidence that soil carbon formation from plant inputs is positively related to
 microbial growth. *Biogeochemistry*, *113 (1)*, 271-281. <u>https://doi.org/10.1007/s10533-012-</u>
 9822-0
- 510 Buckeridge, K. M., Mason, K. E., McNamara, N. P., Ostle, N., Puissant, J., Goodall, T., . . .

- 511 Whitaker, J. (2020). Environmental and microbial controls on microbial necromass recycling,
- an important precursor for soil carbon stabilization. *Communications Earth & Environment, 1*
- 513 (1), 1-9. https://doi.org/10.1038/s43247-020-00031-4
- Camenzind, T., Mason-Jones, K., Mansour, I., Rillig, M. C., & Lehmann, J. (2023). Formation of
 necromass-derived soil organic carbon determined by microbial death pathways. *Nature*

516 *Geoscience, 16 (2)*, 115-122. <u>https://doi.org/10.1038/s41561-022-01100-3</u>

- 517 Chang, Y., Sokol, N. W., van Groenigen, K. J., Bradford, M. A., Ji, D., Crowther, T. W., Liang, C.,
- 518 Luo, Y., Kuzyakov, Y., Wang, J., & Ding, F. (2023). Dataset of manuscript "A stoichiometric
- approach to estimate sources of mineral-associated soil organic matter" [Data set]. Zenodo.
 https://doi.org/10.5281/zenodo.10147884
- 521 Cleveland, C. C., & Liptzin, D. (2007). C: N: P stoichiometry in soil: is there a "Redfield ratio" for 522 the microbial biomass? *Biogeochemistry*, *85 (3)*, 235-252. <u>https://doi.org/10.1007/s10533-</u>
- 523 <u>007-9132-0</u>
- 524 Coonan, E. C., Kirkby, C. A., Kirkegaard, J. A., Amidy, M. R., Strong, C. L., & Richardson, A. E.
- 525 (2020). Microorganisms and nutrient stoichiometry as mediators of soil organic matter
- 526 dynamics. Nutrient Cycling in Agroecosystems, 117, 273-298. https://doi.org/10.1007/s10705-
- 527 <u>020-10076-8</u>
- Cotrufo, M. F., & Lavallee, J. M. (2022). Soil organic matter formation, persistence, and functioning:
 A synthesis of current understanding to inform its conservation and regeneration. *Advances in Agronomy*, *172*, 1-66. <u>https://doi.org/10.1016/bs.agron.2021.11.002</u>
- 531 Cotrufo, M. F., Ranalli, M. G., Haddix, M. L., Six, J., & Lugato, E. (2019). Soil carbon storage 532 informed by particulate and mineral-associated organic matter. *Nature Geoscience, 12 (12)*,

533 989-994. <u>https://doi.org/10.1038/s41561-019-0484-6</u>

- Cotrufo, M. F., Wallenstein, M. D., Boot, C. M., Denef, K., & Paul, E. (2013). The Microbial
 Efficiency-Matrix Stabilization (MEMS) framework integrates plant litter decomposition
 with soil organic matter stabilization: do labile plant inputs form stable soil organic matter? *Global Change Biology, 19 (4)*, 988-995. <u>https://doi.org/10.1111/gcb.12113</u>
- Creamer, C. A., Foster, A. L., Lawrence, C., McFarland, J., Schulz, M., & Waldrop, M. P. (2019).
 Mineralogy dictates the initial mechanism of microbial necromass association. *Geochimica et Cosmochimica Acta*, 260, 161-176. <u>https://doi.org/10.1016/j.gca.2019.06.028</u>
- 541 Creamer, C. A., Jones, D. L., Baldock, J. A., Rui, Y., Murphy, D. V., Hoyle, F. C., & Farrell, M.
- 542 (2016). Is the fate of glucose-derived carbon more strongly driven by nutrient availability,
- soil texture, or microbial biomass size? Soil Biology and Biochemistry, 103, 201-212.
 https://doi.org/10.1016/j.soilbio.2016.08.025
- 545 Crowther, T. W., Hoogen, J. v. d., Wan, J., Mayes, M. A., Keiser, A. D., Mo, L., . . . Maynard, D. S.
- (2019). The global soil community and its influence on biogeochemistry. *Science*, *365*(6455), eaav0550. https://doi.org/10.1126/science.aav0550
- Ding, F., Huang, Y., Sun, W., Jiang, G., & Chen, Y. (2014). Decomposition of organic carbon in fine
 soil particles is likely more sensitive to warming than in coarse particles: an incubation study
 with temperate grassland and forest soils in Northern China. *PLoS One*, 9 (4), e95348.
 https://doi.org/10.1371/journal.pone.0095348
- Ding, F., Sun, W., Huang, Y., & Hu, X. (2018). Larger Q₁₀ of carbon decomposition in finer soil
- 553 particles does not bring long-lasting dependence of Q₁₀ on soil texture. *European Journal of*
- 554 Soil Science, 69 (2), 336-347. <u>https://doi.org/10.1111/ejss.12530</u>

- Ding, X., Liang, C., Zhang, B., Yuan, Y., & Han, X. (2015). Higher rates of manure application lead
 to greater accumulation of both fungal and bacterial residues in macroaggregates of a clay
 soil. *Soil Biology and Biochemistry, 84,* 137-146.
 <u>https://doi.org/10.1016/j.soilbio.2015.02.015</u>
- Elliott, E. T., Anderson, R. V., Coleman, D. C., & Cole, C. V. (1980). Habitable pore space and
 microbial trophic interactions. *Oikos, 35 (3)*, 327-335. <u>https://doi.org/10.2307/3544648</u>
- 561 Fry, B. 2006. Stable Isotope Ecology. New York: Springer. 521, 34-35.
- 562 Georgiou, K., Jackson, R. B., Vindušková, O., Abramoff, R. Z., Ahlström, A., Feng, W., ... Torn, M.
- S. (2022). Global stocks and capacity of mineral-associated soil organic carbon. *Nature Communications, 13 (1)*, 3797. https://doi.org/10.1038/s41467-022-31540-9
- Golchin, A., Oades, J., Skjemstad, J., & Clarke, P. (1994). Study of free and occluded particulate
 organic matter in soils by solid state ¹³C CP/MAS NMR spectroscopy and scanning electron
 microscopy. *Soil Research, 32 (2)*, 285-309. <u>https://doi.org/10.1071/SR9940285</u>
- Grandy, A. S., & Neff, J. C. (2008). Molecular C dynamics downstream: The biochemical
 decomposition sequence and its impact on soil organic matter structure and function. *Science*
- 570 *of The Total Environment, 404 (2),* 297-307. <u>https://doi.org/10.1016/j.scitotenv.2007.11.013</u>
- 571 Griepentrog, M., Bodé, S., Boeckx, P., Hagedorn, F., Heim, A., & Schmidt, M. W. (2014). Nitrogen
- 572 deposition promotes the production of new fungal residues but retards the decomposition of
- old residues in forest soil fractions. *Global Change Biology*, 20 (1), 327-340.
 <u>https://doi.org/10.1111/gcb.12374</u>
- 575 Guigue, J., Just, C., Luo, S., Fogt, M., Schloter, M., Kögel-knabner, I., & Hobley, E. (2021). Spatial 576 molecular heterogeneity of POM during decomposition at different soil depths resolved by

- 577 VNIR hyperspectral imaging. *European Journal of Soil Science*, *73 (1)*, e13207. 578 https://doi.org/10.1111/ejss.13207
- Hu, J., Huang, C., Zhou, S., & Kuzyakov, Y. (2022a). Nitrogen addition to soil affects microbial
 carbon use efficiency: Meta-analysis of similarities and differences in ¹³C and ¹⁸O
 approaches. *Global Change Biology, 28 (16)*, 4977-4988. https://doi.org/10.1111/gcb.16226
- Hu, P., Zhang, W., Chen, H., Xu, L., Xiao, J., Luo, Y., & Wang, K. (2022b). Lithologic control of
 microbial-derived carbon in forest soils. *Soil Biology and Biochemistry*, *167*, 108600.
 https://doi.org/10.1016/j.soilbio.2022.108600
- Islam, M., Singh, B., & Dijkstra, F. A. (2022). Stabilisation of soil organic matter: interactions
 between clay and microbes. *Biogeochemistry*, *160*, 145-158. <u>https://doi.org/10.1007/s10533-</u>
 022-00956-2
- Jasieński, M., & Bazzaz, F. A. (1999). The fallacy of ratios and the testability of models in biology.
 Oikos, 84 (2), 321-326. <u>https://doi.org/10.2307/3546729</u>
- Joergensen, R. G. (2018). Amino sugars as specific indices for fungal and bacterial residues in soil.
 Biology and Fertility of Soils, 54 (5), 559-568. https://doi.org/10.1007/s00374-018-1288-3
- 592 Kaiser, K., Guggenberger, G., & Haumaier, L. (2004). Changes in dissolved lignin-derived phenols,

593 neutral sugars, uronic acids, and amino sugars with depth in forested Haplic Arenosols and

- 594
 Rendzic
 Leptosols.
 Biogeochemistry,
 70
 (1),
 135-151.

 595
 https://doi.org/10.1023/B:BIOG.0000049340.77963.18
- Kaiser, K., Guggenberger, G., Haumaier, L., & Zech, W. (2001). Seasonal variations in the chemical
 composition of dissolved organic matter in organic forest floor layer leachates of old-growth
 Scots pine (*Pinus sylvestris L.*) and European beech (*Fagus sylvatica L.*) stands in

- 599 northeastern Bavaria, Germany. *Biogeochemistry*, 55, 103-143.
 600 https://doi.org/10.1023/A:1010694032121
- Kaiser, K., Guggenberger, G., Haumaier, L., & Zech, W. (2002). The composition of dissolved
 organic matter in forest soil solutions: changes induced by seasons and passage through the
 mineral soil. *Organic Geochemistry*, *33* (*3*), 307-318. <u>https://doi.org/10.1016/S0146-</u>
 <u>6380(01)00162-0</u>
- Kallenbach, C. M., Frey, S. D., & Grandy, A. S. (2016). Direct evidence for microbial-derived soil
 organic matter formation and its ecophysiological controls. *Nature Communications*, 7 (1),

607 13630. <u>https://doi.org/10.1038/ncomms13630</u>

- Kleber, M., Bourg, I. C., Coward, E. K., Hansel, C. M., Myneni, S. C. B., & Nunan, N. (2021).
 Dynamic interactions at the mineral–organic matter interface. *Nature Reviews Earth & Environment, 2 (6)*, 402-421. <u>https://doi.org/10.1038/s43017-021-00162-y</u>
- Klink, S., Keller, A. B., Wild, A. J., Baumert, V. L., Gube, M., Lehndorff, E., . . . Pausch, J. (2022).
 Stable isotopes reveal that fungal residues contribute more to mineral-associated organic
 matter pools than plant residues. *Soil Biology and Biochemistry*, *168*, 108634.
 https://doi.org/10.1016/j.soilbio.2022.108634
- Kögel-Knabner, I. (2017). The macromolecular organic composition of plant and microbial residues
 as inputs to soil organic matter: Fourteen years on. *Soil Biology and Biochemistry*, *105*, A3-
- 617 A8. <u>https://doi.org/10.1016/j.soilbio.2016.08.011</u>
- Kramer, M. G., Sanderman, J., Chadwick, O. A., Chorover, J., & Vitousek, P. M. (2012). Long term
 carbon storage through retention of dissolved aromatic acids by reactive particles in soil. *Global Change Biology, 18 (8),* 2594-2605. <u>https://doi.org/10.1111/j.1365-</u>

621 <u>2486.2012.02681.x</u>

- Lavallee, J. M., Soong, J. L., & Cotrufo, M. F. (2020). Conceptualizing soil organic matter into
 particulate and mineral-associated forms to address global change in the 21st century. *Global Change Biology, 26 (1),* 261-273. <u>https://doi.org/10.1111/gcb.14859</u>
- Lehmann, J., Kinyangi, J., & Solomon, D. (2007). Organic matter stabilization in soil
 microaggregates: implications from spatial heterogeneity of organic carbon contents and
 carbon forms. *Biogeochemistry*, 85 (1), 45-57. <u>https://doi.org/10.1007/s10533-007-9105-3</u>
- Lehmann, J., Solomon, D., Kinyangi, J., Dathe, L., Wirick, S., & Jacobsen, C. (2008). Spatial
- 629 complexity of soil organic matter forms at nanometre scales. *Nature Geoscience, 1 (4)*, 238-
- 630 242. <u>https://doi.org/10.1038/ngeo155</u>
- Leuthold, S. J., Haddix, M. L., Lavallee, J., & Cotrufo, M. F. (2023). Physical fractionation
 techniques. In M. J. Goss & M. Oliver (Eds.), *Encyclopedia of Soils in the Environment (Second Edition)*. Oxford: Academic Press. (Vol. 2, pp. 68-80) <u>https://doi.org/10.1016/B978-</u>
- 634 <u>0-12-822974-3.00067-7</u>
- Liang, C., Amelung, W., Lehmann, J., & Kästner, M. (2019). Quantitative assessment of microbial
 necromass contribution to soil organic matter. *Global Change Biology*, 25 (11), 3578-3590.
 https://doi.org/10.1111/gcb.14781
- Liang, C., & Balser, T. C. (2008). Preferential sequestration of microbial carbon in subsoils of a
 glacial-landscape toposequence, Dane County, WI, USA. *Geoderma*, *148 (1)*, 113-119.
 https://doi.org/10.1016/j.geoderma.2008.09.012
- Liang, C., Kästner, M., & Joergensen, R. G. (2020). Microbial necromass on the rise: The growing
 focus on its role in soil organic matter development. *Soil Biology and Biochemistry*, 150,

643 108000. <u>https://doi.org/10.1016/j.soilbio.2020.108000</u>

- Liang, C., Schimel, J. P., & Jastrow, J. D. (2017). The importance of anabolism in microbial control
 over soil carbon storage. *Nature Microbiology, 2 (8)*, 17105.
 <u>https://doi.org/10.1038/nmicrobiol.2017.105</u>
- Ludwig, M., Achtenhagen, J., Miltner, A., Eckhardt, K.-U., Leinweber, P., Emmerling, C., & Thiele-
- Bruhn, S. (2015). Microbial contribution to SOM quantity and quality in density fractions of
- temperate arable soils. Soil Biology and Biochemistry, 81, 311-322.
 https://doi.org/10.1016/j.soilbio.2014.12.002
- Ma, T., Zhu, S., Wang, Z., Chen, D., Dai, G., Feng, B., . . . Feng, X. (2018). Divergent accumulation
- of microbial necromass and plant lignin components in grassland soils. *Nature Communications*, 9 (1), 3480. <u>https://doi.org/10.1038/s41467-018-05891-1</u>
- Malik, A. A., Puissant, J., Goodall, T., Allison, S. D., & Griffiths, R. I. (2019). Soil microbial
 communities with greater investment in resource acquisition have lower growth yield. *Soil Biology and Biochemistry*, *132*, 36-39. <u>https://doi.org/10.1016/j.soilbio.2019.01.025</u>
- Miltner, A., Bombach, P., Schmidt-Brücken, B., & Kästner, M. (2012). SOM genesis: microbial
 biomass as a significant source. *Biogeochemistry*, 111 (1), 41-55.
 <u>https://doi.org/10.1007/s10533-011-9658-z</u>
- Mortland, M. M. (1959). Reactions of ammonia in soils. *Advances in Agronomy*, 10, 325-348.
 https://doi.org/10.1016/S0065-2113(08)60069-3
- Oldfield, E. E., Crowther, T. W., & Bradford, M. A. (2018). Substrate identity and amount
 overwhelm temperature effects on soil carbon formation. *Soil Biology and Biochemistry*, *124*,
- 664 218-226. <u>https://doi.org/10.1016/j.soilbio.2018.06.014</u>

- 665 Qualls, R. G., & Haines, B. L. (1991). Geochemistry of dissolved organic nutrients in water 666 percolating through a forest ecosystem. *Soil Science Society of America Journal, 55 (4)*,
- 667 1112-1123. https://doi.org/10.2136/sssaj1991.03615995005500040036x
- Ristori, G. (1979). Clay minerals and nutrient availability. *Soils in Mediterranean Type Climates and*

their Yield Potential, 151-163.

- 670 See, C. R., Keller, A. B., Hobbie, S. E., Kennedy, P. G., Weber, P. K., & Pett-Ridge, J. (2022).
- 671 Hyphae move matter and microbes to mineral microsites: Integrating the hyphosphere into
- 672 conceptual models of soil organic matter stabilization. *Global Change Biology, 28 (8)*, 2527-
- 673 2540. <u>https://doi.org/10.1111/gcb.16073</u>
- Simpson, A. J., Simpson, M. J., Smith, E., & Kelleher, B. P. (2007). Microbially derived inputs to
 soil organic matter: are current estimates too low? *Environmental Science & Technology, 41*

676 *(23)*, 8070-8076. <u>https://doi.org/10.1021/es071217x</u>

- Sokol, N. W., Sanderman, J., & Bradford, M. A. (2019). Pathways of mineral-associated soil organic
 matter formation: Integrating the role of plant carbon source, chemistry, and point of entry.
 Global Change Biology, 25 (1), 12-24. https://doi.org/10.1111/gcb.14482
- 680 Sokol, N. W., Slessarev, E., Marschmann, G. L., Nicolas, A., Blazewicz, S. J., Brodie, E. L., . . .
- Consortium, L. S. M. (2022a). Life and death in the soil microbiome: how ecological
 processes influence biogeochemistry. *Nature Reviews Microbiology, 20*, 415-430.
 https://doi.org/10.1038/s41579-022-00695-z
- Sokol, N. W., Whalen, E. D., Jilling, A., Kallenbach, C., Pett-Ridge, J., & Georgiou, K. (2022b).
 Global distribution, formation and fate of mineral-associated soil organic matter under a
 changing climate: A trait-based perspective. *Functional Ecology*, *36* (6), 1411-1429.

687 <u>https://doi.org/10.1111/1365-2435.14040</u>

- 688 Solomon, D., Lehmann, J., Harden, J., Wang, J., Kinyangi, J., Heymann, K., ... Jacobsen, C. (2012).
- 689 Micro- and nano-environments of carbon sequestration: Multi-element STXM–NEXAFS
- 690 spectromicroscopy assessment of microbial carbon and mineral associations. *Chemical*
- 691 *Geology*, *329*, 53-73. <u>https://doi.org/10.1016/j.chemgeo.2012.02.002</u>
- Turrión, M.-B., Glaser, B., & Zech, W. (2002). Effects of deforestation on contents and distribution
 of amino sugars within particle-size fractions of mountain soils. *Biology and Fertility of Soils, 35 (1)*, 49-53. https://doi.org/10.1007/s00374-001-0440-6
- van den Hoogen, J., Geisen, S., Routh, D., Ferris, H., Traunspurger, W., Wardle, D. A., . . . Crowther,
- T. W. (2019). Soil nematode abundance and functional group composition at a global scale.
 Nature, *572 (7768)*, 194-198. <u>https://doi.org/10.1038/s41586-019-1418-6</u>
- Wang, B., An, S., Liang, C., Liu, Y., & Kuzyakov, Y. (2021). Microbial necromass as the source of
 soil organic carbon in global ecosystems. *Soil Biology and Biochemistry*, *162*, 108422.
- 700 <u>https://doi.org/10.1016/j.soilbio.2021.108422</u>
- Wang, X., Wang, C., Cotrufo, M. F., Sun, L., Jiang, P., Liu, Z., & Bai, E. (2020). Elevated
 temperature increases the accumulation of microbial necromass nitrogen in soil via increasing
 microbial turnover. *Global Change Biology, 26 (9),* 5277-5289.
- 704 <u>https://doi.org/10.1111/gcb.15206</u>
- 705 Whalen, E. D., Grandy, A. S., Sokol, N. W., Keiluweit, M., Ernakovich, J., Smith, R. G., & Frey, S.
- 706 D. (2022). Clarifying the evidence for microbial and plant derived soil organic matter, and
- the path towards a more quantitative understanding. *Global Change Biology*, 28 (24), 7167-
- 708 7185. <u>https://doi.org/10.1111/gcb.16413</u>

- Witzgall, K., Vidal, A., Schubert, D. I., Höschen, C., Schweizer, S. A., Buegger, F., . . . Mueller, C.
 W. (2021). Particulate organic matter as a functional soil component for persistent soil
 organic carbon. *Nature Communications, 12 (1)*, 4115. <u>https://doi.org/10.1038/s41467-021-</u>
- 712 <u>24192-8</u>
- Xu, X., Thornton, P. E., & Post, W. M. (2013). A global analysis of soil microbial biomass carbon,
 nitrogen and phosphorus in terrestrial ecosystems. *Global Ecology and Biogeography, 22 (6)*,
 715 737-749. https://doi.org/10.1111/geb.12029
- Yu, W., Huang, W., Weintraub-Leff, S. R., & Hall, S. J. (2022). Where and why do particulate
- organic matter (POM) and mineral-associated organic matter (MAOM) differ among diverse
 soils? Soil Biology and Biochemistry, 172, 108756.
 https://doi.org/10.1016/j.soilbio.2022.108756
- Zhang, Z., Kaye, J. P., Bradley, B. A., Amsili, J. P., & Suseela, V. (2022). Cover crop functional types
 differentially alter the content and composition of soil organic carbon in particulate and
 mineral-associated fractions. *Global Change Biology*, 28 (19), 5831-5848.
 https://doi.org/10.1111/gcb.16296
- 724
- 725
- 726
- 727
- 728 Figure legends

FIGURE 1 Schematic diagram to illustrate the two-end-member mixing model to estimate the
 contribution of microbial and plant residues to mineral-associated organic matter (MAOM).

Fractional abundance of N [N/(C+N)] in particulate organic matter (POM) and microbial biomass
were used as end-members in a mixing model to estimate plant versus microbial contributions to
MAOM. As an illustrative example, the figure shows a scenario when C/N ratios are 20, 15 and 10
for POM, MAOM and microbes, respectively.

735

FIGURE 2 Comparison of C/N ratios of particulate organic matter (POM), mineral-associated organic matter (MAOM), and microbial biomass (microbe) across all ecosystems (N = 288) (a), and in forests (N = 64) (b), grasslands (N = 69) (c), and croplands (N = 146) (d). The plots display the individual C/N ratios, as well as their density distribution, means and standard deviations. *** indicates p < 0.001, * indicates p < 0.05.

741

FIGURE 3 Regressions between the C/N ratios of mineral-associated organic matter (C/N_{MAOM}) and microorganisms (C/N_{Microbe}), or particulate organic matter (C/N_{POM}) across all ecosystems (a) and separately for forests (b), grasslands (c), and croplands (d). The X-axis refers to the C/N ratio of microbial biomass or POM. The black dashed line shows the 1:1 relationship. A few data points representing C/N ratios of POM fall outside the range of the X-axis (> 40), as do a few MAOM data points for the Y-axis (> 25), and consequently are not shown here (see Table S2).

748

FIGURE 4 Microbial contribution to MAOM depends on ecosystem type (a) and soil depth (b).
Abnormal points (with contribution equal to 0 or 100%) were excluded in this analysis. The box
represents the upper and lower quartiles. The top and bottom whisker-line represent the maximum
and minimum values, the black dot and nearby black number in the box represents the mean value,

and the horizontal line in the box represents the median. The scattered points within the box and
whisker-line display the individual contributions. The numbers in the parentheses indicate the sample
size.

756

757 FIGURE 5 Relative importance of climate and soil properties to predict microbial contribution

to MAOM by random forest analysis. The abnormal points (with contribution equal to 0 or 100%) were excluded in this analysis. Red bars indicate that the factor is positively correlated with the microbial contribution to MAOM; blue bars indicate that the factor is negatively correlated with the microbial contribution to MAOM. %IncMSE, percentage of increase of mean square error (%). MAP: mean annual precipitation, MAT: mean annual temperature.

763



764 765

FIGURE 1







Quantify the sources of MAOM by stoichiometry





